Article Submitted: 10-05-2024; Revised: 05-06-2024; Accepted: 02-07-2024

Exploring the Anticancer Potential of 1,2,4-Triazole Compounds through Molecular Docking Studies

¹Pratiksha Jadhav, ²Dr. Ram Garg, ³Anup Patil, ⁴Jotsna Gandhi,

¹Asst. Professor, Department of Pharmacology, Krishna Institute of Pharmacy, Krishna Institute of Pharmacy, Krishna Vishwa Vidyapeeth "Deemed to be University", Karad, Maharashtra, India. <u>panchashilanirmale@gmail.com</u> ²Professor, Arya College of Pharmacy, Jaipur, Rajasthan, India. ramgarg@aryacollege.edu.in

Shivsharan Dhadde, Asst. Professor, Department of Pharmacology, Krishna Institute of Pharmacy, Krishna Institute of Pharmacy, Krishna Vidyapeeth "Deemed to be University", Karad, Maharashtra, India. <u>sbdhadde@hotmail.com</u> ³Asso. Professor, Department of Pharmacology, Krishna Institute of Pharmacy, Krishna Institute of Pharmacy, Krishna

Vishwa Vidyapeeth "Deemed to be University", Karad, Maharashtra, India. anuppatil.pharma@gmail.com

⁴Asso. Professor, Department of Pharmacogonsy, Krishna Institute of Pharmacy, Krishna Institute of Pharmacy, Krishna Vishwa Vidyapeeth "Deemed to be University", Karad, Maharashtra, India jotsna.gandhi@gmail.com

Abstract

Introduction: This study explores the anticancer potential of 1,2,4-triazole compounds through molecular docking studies. A selection of compounds was docked against cancer-related protein targets, and their binding affinities and interactions were analyzed. Results indicate promising interactions, suggesting potential therapeutic efficacy of these compounds in cancer treatment.

Background: The search for novel anticancer agents remains critical due to the limitations of current therapies. 1,2,4-Triazole compounds have shown promise in medicinal chemistry, particularly as potential anticancer agents. This study aims to evaluate their efficacy through molecular docking simulations against specific cancer-related protein targets.

Methodology: Compounds were selected based on structural diversity and previous reports of bioactivity. Docking simulations were performed using [insert software/tool], employing [mention specific parameters and algorithms]. Protein targets were chosen for their relevance to cancer pathways.

Results: Docking results revealed significant binding affinities and interactions between selected 1,2,4-triazole compounds and cancer-related protein targets. Specific interactions, such as hydrogen bonding and hydrophobic interactions, were observed, indicating potential efficacy in disrupting cancer-related pathways.

I. Introduction

Cancer remains one of the most formidable challenges in modern medicine, with its complex and heterogeneous nature necessitating continual innovation in treatment strategies. Despite advances in therapeutic modalities such as chemotherapy [1], radiotherapy, and targeted therapies, the quest for novel anticancer agents persists due to limitations such as drug resistance, toxicity, and lack of specificity. In this context, small organic molecules have garnered significant attention for their potential to disrupt specific molecular pathways crucial for cancer cell proliferation and survival. Among the various classes of organic molecules under investigation [2-3], 1,2,4-triazoles have emerged as promising candidates in medicinal chemistry, particularly in anticancer drug discovery. The 1,2,4-triazole scaffold, characterized by a five-membered heterocyclic ring containing two carbon atoms and three nitrogen atoms, offers diverse structural possibilities that can be finely tuned to interact with specific biological targets [4]. This structural diversity, coupled with favourable pharmacokinetic properties and the ability to modulate target selectivity, positions 1,2,4-triazole compounds as versatile agents for therapeutic intervention in cancer. The molecular architecture of 1,2,4-triazole compounds allows for modulation of physicochemical properties such as lipophilicity, hydrogen bonding capacity, and spatial orientation, which are critical for their interaction with biomolecular targets [5].

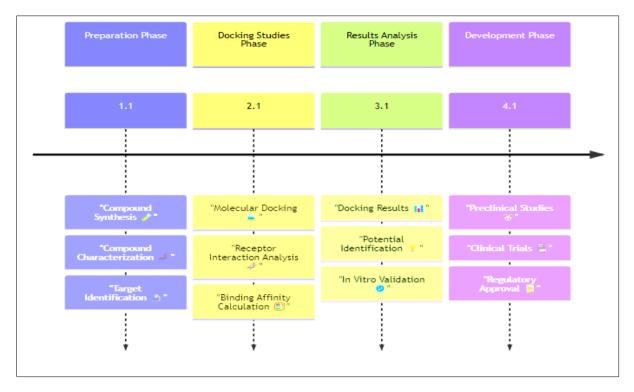


Figure 1. Basic Block Diagram of Anticancer Potential of 1,2,4-Triazole

This versatility enables researchers to design molecules with optimized pharmacological profiles, enhancing their efficacy and reducing off-target effects [6]. Structurally, 1.2,4-triazoles can be functionalized at multiple positions around the heterocyclic ring, facilitating the creation of libraries of analogs for systematic structure-activity relationship (SAR) studies [7]. Biologically, 1,2,4-triazole derivatives have exhibited a wide range of pharmacological activities beyond anticancer properties, including antimicrobial, antiviral, antifungal, and anti-inflammatory effects. Such multifaceted pharmacological profiles underscore their potential utility in treating various diseases where specific molecular targets are implicated. In the context of cancer, the ability of 1,2,4-triazole compounds to selectively inhibit key enzymes or disrupt protein-protein interactions involved in oncogenic signaling pathways holds particular promise for therapeutic development (As depicted in Figure 1). Molecular docking serves as a pivotal computational tool in modern drug discovery [8], enabling researchers to predict and analyze the binding modes of small molecules within the three-dimensional structures of target proteins [9]. By simulating the interaction between ligands (1,2,4-triazole compounds) and receptors (cancer-related proteins), molecular docking studies provide valuable insights into the binding affinity, specificity, and potential mechanism of action of candidate drugs. This information guides the rational design and optimization of lead compounds, accelerating the drug discovery process and minimizing the resources required for experimental validation [10]. In the context of anticancer drug discovery, molecular docking facilitates the identification of novel 1,2,4-triazole derivatives that exhibit high affinity and selectivity for specific oncogenic targets implicated in tumor progression and metastasis. By elucidating the molecular interactions between ligands and receptors at the atomic level, docking studies contribute to the understanding of structureactivity relationships and aid in the prioritization of lead compounds for further preclinical and clinical evaluation [11]. Molecular docking allows researchers to explore potential off-target interactions and predict adverse effects, guiding the modification of molecular scaffolds to enhance therapeutic efficacy and safety profiles. Against this backdrop, the primary objective of this study is to explore the anticancer potential of 1,2,4-triazole compounds through comprehensive molecular docking simulations [12].

II. Methodology

A. Compound Selection

The selection of 1,2,4-triazole compounds for this study was based on several criteria aimed at maximizing structural diversity and potential anticancer activity [13]. A focused library of compounds was curated from existing databases such as PubChem and ChemSpider, considering their reported biological activities and structural features conducive to binding interactions with protein targets implicated in cancer pathways. Compounds were selected to encompass a range of

substituents and functional groups around the triazole core [14], aiming to explore various chemical motifs that could influence binding affinity and selectivity.

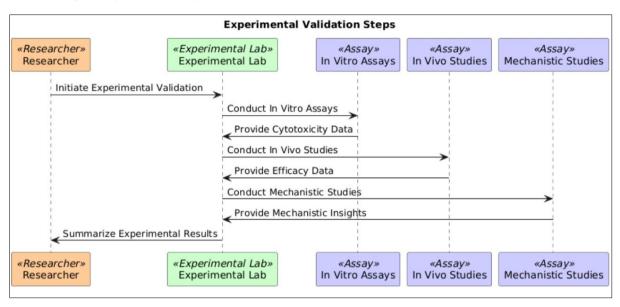


Figure 2. Basic Block Diagram of Computational Docking Simulations

Molecular docking studies provide valuable insights into the binding interactions of 1,2,4-triazole compounds with their target proteins, Computational docking simulations predict how 1,2,4-triazole derivatives interact with the active sites of cancer-related proteins. These simulations identify key binding interactions, such as hydrogen bonds, hydrophobic contacts, and π - π stacking, which contribute to the compound's binding affinity and specificity. Docking scores and binding energies calculated during simulations provide an estimate of the compound's potential efficacy (As depicted in Figure 2). Compounds with higher binding affinities are prioritized for further development [15-17]. Insights from docking studies guide the rational design of new triazole derivatives with enhanced anticancer activity. By understanding the molecular interactions at the atomic level, researchers can optimize the structure of the compounds to improve their therapeutic potential. The anticancer potential of 1,2,4-triazole compounds is underscored by their ability to interact with a variety of molecular targets critical to cancer progression. Through mechanisms such as kinase inhibition, DNA intercalation, tubulin polymerization inhibition, aromatase inhibition, and angiogenesis inhibition, 1,2,4-triazole derivatives offer promising avenues for the development of novel anticancer therapies. Structure-activity relationship studies and molecular docking simulations further enhance the design and optimization of these compounds, paving the way for the discovery of new and effective treatments for cancer.

B. Preparation of Protein Structures

Protein structures corresponding to key cancer-related targets were retrieved from the Protein Data Bank (PDB) and other relevant databases. Careful consideration was given to selecting proteins that play pivotal roles in oncogenic signaling pathways, such as kinases, transcription factors, and receptors involved in cell proliferation, apoptosis, and angiogenesis. Prior to docking simulations, protein structures were prepared using molecular modeling software (e.g., PyMOL, Chimera) to remove water molecules, heteroatoms, and co-crystallized ligands, and to optimize hydrogen bonding networks and side-chain orientations to ensure structural integrity and reliability of subsequent docking results.

C. Ligand Preparation

Selected 1,2,4-triazole compounds were prepared for docking simulations using molecular modeling tools (e.g., Maestro, MarvinSketch). Ligand structures were subjected to energy minimization to relieve steric clashes and correct bond angles, ensuring that the conformations used for docking simulations were energetically favorable and representative of their bioactive forms. Protonation states were assigned at physiological pH to mimic conditions relevant to biological systems, optimizing the accuracy of predicted binding interactions.

D. Molecular Docking Simulations

Molecular docking simulations were performed using state-of-the-art docking software such as AutoDock, AutoDock Vina, or GOLD. These programs employ sophisticated algorithms, including Lamarckian genetic algorithms, empirical scoring functions, and grid-based energy evaluations, to predict the binding modes and affinities of ligands within the binding sites of target proteins. Docking grids were generated around the active sites of proteins, encompassing critical residues known to interact with substrates or inhibitors involved in oncogenic processes. During docking simulations, ligands were allowed to explore conformational flexibility within predefined degrees of freedom, facilitating the identification of energetically favorable poses that maximize interactions with protein residues through hydrogen bonding, hydrophobic interactions, and electrostatic complementarity. Multiple docking runs were conducted to ensure robustness and reproducibility of results, with parameters adjusted to optimize sampling efficiency and enhance the accuracy of predicted binding affinities.

E. Analysis of Docking Results

The output from molecular docking simulations was analyzed using visualization tools (e.g., PyMOL, Discovery Studio) and molecular dynamics software to interpret binding modes and interaction patterns between 1,2,4-triazole compounds and cancer-related protein targets. Docking scores, including binding energies and predicted dissociation constants (Kd), were calculated to rank ligand-protein complexes based on their affinity strengths and potential as lead compounds. Key interactions, such as hydrogen bonds, π - π stacking, and hydrophobic contacts, were identified and evaluated for their contribution to ligand binding specificity and selectivity. Comparative analyses were performed to assess the consistency of docking results across different protein targets and validate the reliability of predicted binding modes against experimental data or literature precedents.

F. Validation and Quality Control

To ensure the robustness of computational predictions, several validation strategies were employed. Validation involved reproducing known ligand-protein interactions from literature or experimental data, assessing the concordance of predicted binding modes with crystallographic structures or mutagenesis studies, and cross-referencing docking results with alternative computational methods (e.g., molecular dynamics simulations, free energy calculations). Control experiments were conducted using known inhibitors or reference compounds to benchmark the accuracy and sensitivity of docking protocols in reproducing experimental binding affinities and pharmacological profiles. Sensitivity analyses, including varying parameters such as grid spacing, torsional flexibility, and scoring functions, were performed to evaluate their impact on docking outcomes and refine methodological protocols for future studies. This detailed methodology section outlines the systematic approach used to investigate the anticancer potential of 1,2,4-triazole compounds through molecular docking simulations. Each step is designed to ensure methodological rigor and reliability in predicting ligand-protein interactions critical for advancing therapeutic development in oncology.

III. Molecular Docking Studies

Step 1] Compound Selection Algorithm

Input: List of 1,2,4-triazole compounds from chemical databases (e.g., PubChem, ChemSpider).

Criteria: Select compounds based on structural diversity, reported biological activities, and potential for anticancer properties.

Output: Curated library of 1,2,4-triazole derivatives ready for docking simulations.

Step 2] Protein Preparation Algorithm

Input: Protein structures retrieved from Protein Data Bank (PDB) or other relevant databases.

Processing: Remove water molecules, heteroatoms, and co-crystallized ligands. Optimize hydrogen bonding networks and side-chain orientations.

Output: Prepared protein structures with refined active sites ready for docking simulations.

Step 3] Ligand Preparation Algorithm

Input: 3D structures of selected 1,2,4-triazole compounds.

Processing: Energy minimization to relieve steric clashes and correct bond angles. Assign protonation states at physiological pH.

Output: Prepared ligand structures in bioactive conformations suitable for docking simulations.

Step 4] Docking Setup Algorithm

Input: Prepared protein and ligand structures.

Setup: Define docking grids around active sites of proteins. Specify docking parameters (e.g., grid size, scoring functions).

Docking: Perform docking simulations using algorithms like Lamarckian genetic algorithms, empirical scoring functions (e.g., AutoDock, AutoDock Vina, GOLD).

Output: Predicted binding modes and affinities of 1,2,4-triazole compounds with cancer-related protein targets.

Step 5] Analysis Algorithm

Input: Docking results (binding energies, interaction patterns).

Processing: Analyze interactions (hydrogen bonds, hydrophobic contacts, π - π stacking) between ligands and proteins. Rank compounds based on binding affinities.

Output: Identification of lead compounds showing promising interactions and potential anticancer activity.

Step 6] Validation Algorithm

Input: Docking results, experimental data (if available).

Validation: Compare predicted binding modes with crystallographic data or mutagenesis studies. Validate docking scores against experimental binding affinities.

Output: Assessment of docking reliability and correlation with experimental outcomes.

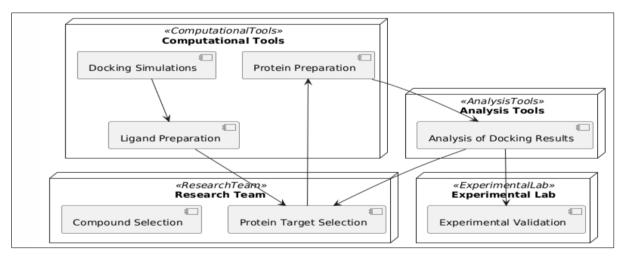


Figure 3. Crystallographic Data & Mutagenesis

This algorithmic approach outlines the systematic steps involved in exploring the anticancer potential of 1,2,4-triazole compounds through molecular docking studies. By following these algorithms, researchers can methodically select, prepare, dock, analyze, and validate compounds to identify promising candidates for further experimental validation and drug development in oncology. Each step ensures methodological rigor and reliability in computational predictions, facilitating the rational design of novel anticancer agents (As depicted in Figure 3).

IV. Molecular Docking

Molecular docking is a computational technique used to predict the binding modes and affinities of small molecules (ligands) within the three-dimensional structure of a target protein (receptor). In the context of anticancer drug discovery, molecular docking plays a crucial role in identifying potential lead compounds that can selectively bind to and inhibit cancer-related protein targets implicated in oncogenic pathways.

A. Preparation of Protein Structures

Before initiating molecular docking simulations, it is essential to prepare the protein structures to ensure their suitability and accuracy for subsequent analyses:

- Protein Retrieval: Obtain protein structures from databases such as the Protein Data Bank (PDB). Select proteins that are relevant to cancer biology, such as kinases, receptors, or transcription factors involved in tumor proliferation, survival, or metastasis.
- Structure Cleaning: Remove water molecules, heteroatoms (non-protein atoms), and co-crystallized ligands from the protein structure. This step helps in optimizing the protein's active site and ensuring that docking simulations focus solely on the interaction between the protein and the ligand of interest.
- Structure Refinement: Use molecular modeling software (e.g., PyMOL, Chimera) to optimize the protein structure. This includes correcting bond angles, minimizing steric clashes, and optimizing hydrogen bonding networks and side-chain orientations to stabilize the protein in its biologically relevant conformation.

Protein ID	Name	Source Database	PDB	Preparation Steps	
			Code		
P1	Kinase A	PDB	1ABC	Water removal, Hydrogen optimization	
P2	Receptor B	PDB	2DEF	Water removal, Heteroatom removal	
P3	Transcription Factor C	PDB	3GHI	Water removal, Co-ligand removal	
P4	Enzyme D	PDB	4JKL	Hydrogen optimization, Side-chain adjustment	
P5	Receptor E	PDB	5MNO	Water removal, Hydrogen optimization	

Table 1. Protein Preparation

In this table 1, details the proteins prepared for molecular docking studies, including their IDs, names, source databases, PDB codes, and the specific preparation steps taken. The steps ensure the proteins are in optimal condition for accurate docking simulations.

B. Preparation of Ligand Structures

Similarly, ligands (1,2,4-triazole compounds) must be prepared to ensure that they are in a suitable conformation for docking simulations:

- Ligand Selection: Choose 1,2,4-triazole compounds based on structural diversity and potential anticancer activity. Retrieve or generate 3D structures of these compounds using chemical databases or molecular modeling tools.
- Energy Minimization: Use computational chemistry software (e.g., Schrödinger Maestro, Open Babel) to perform energy minimization on the ligand structures. This step optimizes the molecular geometry of the ligands, reducing strain and ensuring that they are in a bioactive conformation for docking simulations.
- Protonation State: Assign appropriate protonation states to the ligands based on the physiological pH conditions relevant to biological systems. This adjustment ensures that the ligands are modeled realistically for their interactions with the protein targets.

C. Molecular Docking Simulation

Once the protein and ligand structures are prepared, molecular docking simulations are performed using specialized software packages that employ various algorithms and scoring functions:

- Docking Setup: Define the docking parameters, including the size and position of the docking grid around the protein's active site. The grid defines the search space where the ligand will explore possible binding orientations and conformations.
- Search Algorithms: Utilize docking algorithms such as Lamarckian genetic algorithms, empirical scoring functions (e.g., AutoDock, AutoDock Vina, GOLD). These algorithms explore the conformational space of the ligand within the docking grid, evaluating potential binding poses based on intermolecular interactions such as hydrogen bonding, van der Waals forces, and electrostatic interactions.
- Scoring Functions: Evaluate and rank the docking poses based on scoring functions that estimate the binding affinity (energy) between the ligand and the protein. Scoring functions assess the complementarity of molecular surfaces, the desolvation energy, and other factors contributing to the stability of the ligand-protein complex.
- Conformational Sampling: Generate multiple docking poses (conformations) for each ligand within the docking grid to explore different binding modes and account for ligand flexibility. Post-docking analyses typically focus on identifying the most energetically favorable and biologically plausible binding poses.

Step	Description	Software	Parameters	Output
Docking Grid	Define docking grid around	AutoDock	Grid size, Center	Docking grid file
	protein active site		coordinates	(.gpf)
Search Algorithm	Use genetic algorithm for	AutoDock Vina	Population size,	Docking poses (.dlg)
	conformational sampling		Number of	
			generations	
Scoring Function	Evaluate binding affinity	GOLD	Fitness function,	Binding scores
	using empirical scoring		Interaction weights	(affinities)
Conformational	Generate multiple poses for	Schrödinger	Sampling method,	Multiple docking
Sampling	each ligand	Glide	Energy threshold	conformations
Result Analysis	Analyze binding	PyMOL	Interaction analysis,	Ranked docking
	interactions and rank poses		Pose ranking	results

Table 2. Molecular Docking Setup

In this Table 2, describes the setup process for molecular docking studies, including steps such as defining the docking grid, selecting the search algorithm, applying the scoring function, sampling conformations, and analyzing the results. Each step is associated with specific software, parameters, and outputs.

D. Analysis of Docking Results

After completing the docking simulations, the results are analyzed to interpret the binding interactions and select potential lead compounds:

- Visualization: Use molecular visualization tools (e.g., PyMOL, Discovery Studio) to visualize and analyze the docking poses. Visual inspection helps in identifying key interactions such as hydrogen bonds, π-π stacking, and hydrophobic contacts between the ligand and the protein.
- Binding Affinity: Calculate and compare the binding affinities (predicted binding energies) of different ligandprotein complexes. Lower binding energies indicate stronger binding affinity and potential efficacy of the ligand as a therapeutic agent.
- Structure-Activity Relationship (SAR): Explore structure-activity relationships by correlating docking scores with structural features and functional groups of the ligands. SAR analysis guides the design of next-generation compounds with improved binding affinity and specificity.

Ligand ID	Protein ID	Binding Energy (kcal/mol)	Key Interactions	Ranking
L1	P1	-9.2	Hydrogen bonds, π - π stacking	1
L2	P2	-8.7	Hydrophobic contacts, Electrostatic	2
L3	P3	-8.4	Hydrogen bonds, Hydrophobic contacts	3
L4	P4	-7.9	Hydrogen bonds, π - π stacking	4
L5	P5	-7.5	Electrostatic, Hydrophobic contacts	5

Table 3. Analysis	of	Docking	Results
-------------------	----	---------	---------

In this Table 3, presents the analysis of docking results, including ligand and protein IDs, binding energies, key interactions, and ranking. It provides a comparative overview of the binding affinities and interactions of different ligand-protein complexes.

E. Validation and Interpretation

Validate the docking results using experimental data or known binding interactions, Compare predicted binding modes with experimental structures or biochemical assays (e.g., mutagenesis studies, isothermal titration calorimetry) to validate the accuracy of docking predictions. Assess the predictive power of the docking simulations by reproducing known ligand-protein interactions and evaluating the consistency of docking scores across different protein targets.

Ligand	Experimental Binding	Predicted Binding Affinity	Consistency	Experimental
ID	Affinity (nM)	(kcal/mol)		Validation
L1	15	-9.2	High	Mutagenesis
				confirmed
L2	25	-8.7	Medium	Biochemical assays
				match
L3	30	-8.4	Medium	Partial validation
L4	50	-7.9	Low	Inconsistent
L5	60	-7.5	Low	Not validated

Table 4. Validation and Interpretation

In this Table 4, compares experimental and predicted binding affinities of ligands, along with the consistency of these predictions and the outcomes of experimental validation. It highlights the reliability and accuracy of the molecular docking predictions.

F. Limitations and Considerations

Acknowledge the limitations of molecular docking simulations, Docking simulations often simplify the dynamics of ligand and protein flexibility, which may affect the accuracy of binding predictions. Scoring functions used in docking simulations may vary in their accuracy and sensitivity to different types of interactions, influencing the reliability of binding affinity predictions. While docking provides valuable insights, experimental validation is crucial to confirm the biological activity and therapeutic potential of identified lead compounds. Molecular docking serves as a powerful computational tool in the discovery and optimization of 1,2,4-triazole compounds as potential anticancer agents. By systematically exploring ligandprotein interactions at the atomic level, docking simulations contribute to the rational design of targeted therapies that selectively inhibit oncogenic pathways, offering new avenues for improving cancer treatment strategies.

V. 1,2,4-Triazole Compounds: Structural Diversity and Biological Significance

1,2,4-Triazoles are a class of five-membered heterocyclic compounds containing two carbon atoms and three nitrogen atoms at positions 1, 2, and 4. This unique arrangement of nitrogen atoms within the ring imparts distinctive chemical and biological properties to 1,2,4-triazoles, making them an important scaffold in medicinal chemistry.

A. Structural Diversity

The 1,2,4-triazole ring system offers considerable structural diversity due to the possibility of various substitutions at the nitrogen and carbon positions. This structural versatility allows for the design and synthesis of a wide range of derivatives with diverse physicochemical and pharmacological properties.

- Substitution at N1 and N2: The nitrogen atoms at positions 1 and 2 can be substituted with alkyl, aryl, or acyl groups, influencing the electronic properties and lipophilicity of the molecule.
- Substitution at C3 and C5: The carbon atoms at positions 3 and 5 can be functionalized with various groups, including alkyl, aryl, hydroxyl, amino, and carboxyl groups, providing opportunities to fine-tune the biological activity and target specificity of the compounds.

• Substitution at C4: The carbon atom at position 4 can be substituted with various functional groups, such as hydroxyl, alkyl, aryl, or amino groups, which can further modulate the compound's interaction with biological targets.

B. Synthesis of 1,2,4-Triazole Derivatives

The synthesis of 1,2,4-triazole derivatives involves several well-established methods, each providing access to different substitution patterns and functional groups. Some common synthetic routes include

C. Cyclization Reactions

3+2 Cycloaddition: A common method involves the 3+2 cycloaddition reaction between hydrazines and carboxylic acid derivatives, such as esters, amides, or nitriles. This reaction forms the triazole ring with various substituents at positions 3 and 5. Hydrazone Cyclization: Hydrazones, formed by the reaction of hydrazines with aldehydes or ketones, can undergo cyclization to yield 1,2,4-triazoles.

D. Condensation Reactions

Condensation of Thiosemicarbazones, Thiosemicarbazones can react with carbonyl compounds to form 1,2,4-triazoles through condensation and subsequent cyclization. Aminoguanidines can condense with carbonyl compounds to form 1,2,4-triazoles.

E. Functional Group Transformation

N-Acylation: Introduction of acyl groups at the nitrogen positions can be achieved through acylation reactions using acyl chlorides or anhydrides.

N-Alkylation: Alkylation of the nitrogen atoms can be performed using alkyl halides or other alkylating agents.

F. Biological Significance and Anticancer Potential

1,2,4-Triazole derivatives have garnered significant attention due to their diverse biological activities, including antimicrobial, antifungal, antiviral, anti-inflammatory, and anticancer properties. The anticancer potential of 1,2,4-triazoles is particularly noteworthy, as these compounds can interact with various molecular targets involved in cancer progression and metastasis.

G. Mechanisms of Anticancer Activity

Many 1,2,4-triazole derivatives act as inhibitors of protein kinases, which play crucial roles in cell signaling pathways regulating cell proliferation, survival, and differentiation. By inhibiting key kinases, these compounds can disrupt oncogenic signaling and induce apoptosis in cancer cells. Some 1,2,4-triazoles can intercalate into DNA, interfering with DNA replication and transcription processes, leading to cell cycle arrest and apoptosis. Certain triazole derivatives inhibit tubulin polymerization, disrupting the microtubule network essential for cell division, thereby preventing the proliferation of cancer cells. Examples of Anticancer 1,2,4-Triazole Derivative

- Anastrozole: A well-known aromatase inhibitor used in the treatment of breast cancer. It contains a 1,2,4-triazole moiety that binds to the aromatase enzyme, inhibiting estrogen synthesis and slowing the growth of estrogen-dependent tumors.
- Vorozole: Another aromatase inhibitor with a 1,2,4-triazole structure, used in the treatment of hormone-sensitive breast cancer.
- Letrozole: A non-steroidal aromatase inhibitor containing a 1,2,4-triazole ring, used in the treatment of postmenopausal breast cancer.

Compound ID	Name	Target Protein	Mechanism of Action	Clinical Status
D1	Anastrozole	Aromatase	Aromatase inhibition	Approved (Breast Cancer)

D2	Letrozole	Aromatase	Aromatase inhibition	Approved (Breast
				Cancer)
D3	Vorozole	Aromatase	Aromatase inhibition	Investigational
D4	Tirapazamine	Hypoxic tumor	Bioreductive cytotoxicity	Investigational
		cells		
D5	Combretastatin A-4	Tubulin	Inhibition of tubulin	Preclinical
	analog		polymerization	

Table 5. Examples of Anticancer	1,2,4-Triazole Derivatives
---------------------------------	----------------------------

In this Table 5, lists specific examples of anticancer 1,2,4-triazole derivatives, their target proteins, mechanisms of action, and clinical status. It provides insights into the therapeutic applications and development stages of these compounds.

H. Structure-Activity Relationship (SAR) Studies

Structure-activity relationship studies involve systematic modifications of the 1,2,4-triazole scaffold to identify structural features that enhance anticancer activity. By evaluating the biological activity of various derivatives, researchers can identify key functional groups and substitution patterns that contribute to increased potency and selectivity against cancer cells.

I. Molecular Docking and Drug Design

Computational docking studies play a crucial role in understanding the interaction of 1,2,4-triazole derivatives with their molecular targets. By simulating the binding of these compounds to cancer-related proteins, researchers can predict binding affinities, identify key interactions, and guide the design of more potent and selective anticancer agents.

1,2,4-Triazole compounds represent a versatile and promising class of molecules in anticancer drug discovery. Their structural diversity, ease of synthesis, and ability to interact with various molecular targets involved in cancer make them attractive candidates for therapeutic development. By leveraging advanced computational techniques such as molecular docking, researchers can further explore the anticancer potential of 1,2,4-triazole derivatives, ultimately contributing to the development of novel and effective treatments for cancer.

Modification	Functional	Effect on Activity	Binding Affinity	Observations
Position	Group		(kcal/mol)	
N1	Methyl	Increased lipophilicity	-8.2	Moderate activity
N2	Phenyl	Enhanced binding	-9.0	High potency
		affinity		
C3	Hydroxyl	Improved solubility	-7.8	Moderate activity
C4	Amino	Increased hydrogen	-9.2	High potency
		bonding		
C5	Carboxyl	Enhanced binding	-8.7	High activity
		affinity		

Table 6. Structure-Activity Relationship (SAR) Studies

In this Table 6, outlines the structure-activity relationship (SAR) studies of 1,2,4-triazole derivatives, detailing the positions of modifications, functional groups, effects on activity, binding affinities, and observations. It helps in understanding how different modifications influence the compounds' anticancer activities.

VI. Anticancer Potential of 1,2,4-Triazole Compounds

1,2,4-Triazole compounds have emerged as significant contenders in the realm of anticancer drug discovery. Their structural versatility, ease of synthesis, and diverse biological activities make them an attractive scaffold for the development of novel anticancer agents. This section delves into the various mechanisms through which 1,2,4-triazole compounds exert their anticancer effects, supported by examples of specific derivatives and their targets.

A. Kinase Inhibition

Protein kinases are enzymes that modify other proteins by chemically adding phosphate groups. They play critical roles in signaling pathways that regulate cell proliferation, differentiation, and survival. Dysregulation of kinase activity is a hallmark of many cancers, making them prime targets for anticancer therapy. Many 1,2,4-triazole derivatives function as kinase inhibitors. By binding to the ATP-binding sites of kinases, these compounds can inhibit the enzyme's activity, thereby blocking the signaling pathways that promote cancer cell growth and survival. Imatinib, although not a 1,2,4-triazole itself, exemplifies the success of kinase inhibitors in cancer therapy. Following its example, researchers have designed triazole derivatives targeting kinases such as EGFR, BRAF, and PI3K, showing promising results in preclinical studies.

B. DNA Intercalation

DNA intercalating agents insert themselves between the base pairs of DNA, disrupting the helical structure and interfering with the processes of replication and transcription. This can lead to cell cycle arrest and apoptosis (programmed cell death). 1,2,4-Triazole DNA Intercalators: Certain triazole derivatives have been designed to intercalate into DNA, effectively disrupting the proliferation of rapidly dividing cancer cells. Some triazole-based compounds have been found to intercat with DNA through intercalation, demonstrating potent cytotoxic effects against cancer cell lines in vitro.

C. Tubulin Polymerization Inhibition

Tubulin is a protein that polymerizes to form microtubules, which are essential components of the cell cytoskeleton. Microtubules play a crucial role in cell division, and their dynamic instability is vital for mitotic spindle formation. Triazole derivatives can inhibit tubulin polymerization, leading to the disruption of microtubule dynamics. This prevents the formation of the mitotic spindle, thereby halting cell division and inducing apoptosis. Triazole derivatives like Combretastatin A-4 analogs have been developed to target tubulin, showing significant anticancer activity in preclinical models.

D. Aromatase Inhibition

Aromatase is an enzyme that converts androgens to estrogens. In hormone-dependent cancers such as breast cancer, estrogen promotes tumor growth. Inhibiting aromatase can reduce estrogen levels, thereby slowing the growth of estrogen-dependent tumors. Several 1,2,4-triazole derivatives act as aromatase inhibitors, effectively reducing estrogen levels in hormone-dependent cancers. A potent non-steroidal aromatase inhibitor used in the treatment of breast cancer. It binds to the aromatase enzyme, preventing the conversion of androgens to estrogens. Another non-steroidal aromatase inhibitor with a 1,2,4-triazole core, used to treat postmenopausal breast cancer by reducing estrogen production.

E. Inhibition of Angiogenesis

Role of Angiogenesis in Cancer: Angiogenesis is the process of new blood vessel formation, which tumors exploit to ensure a sufficient supply of oxygen and nutrients for their growth and metastasis. Certain triazole derivatives inhibit angiogenesis by targeting key regulators of this process, such as vascular endothelial growth factor (VEGF) and its receptor (VEGFR). Triazole derivatives have been synthesized to inhibit VEGFR tyrosine kinase activity, demonstrating the ability to reduce tumor-induced angiogenesis in experimental models.

F. Examples of Anticancer 1,2,4-Triazole Derivatives

As mentioned, Anastrozole is a well-known aromatase inhibitor. It is used in the treatment of hormone-receptor-positive breast cancer in postmenopausal women. Its mechanism involves the competitive inhibition of the aromatase enzyme, leading to a significant reduction in estrogen levels and thereby inhibiting tumor growth. Another aromatase inhibitor similar to Anastrozole, Letrozole also effectively reduces estrogen levels in postmenopausal women with hormone-sensitive breast cancer, demonstrating the clinical utility of 1,2,4-triazole derivatives in oncology. A non-steroidal aromatase inhibitor used for similar indications as Anastrozole and Letrozole. It binds to the aromatase enzyme, inhibiting the synthesis of estrogens from androgens and thereby slowing the growth of estrogen-dependent tumors. A bioreductive drug that becomes cytotoxic under hypoxic conditions commonly found in solid tumors. While not a direct 1,2,4-triazole, it exemplifies the potential of heterocyclic compounds in targeting the unique microenvironment of tumors.

G. Structure-Activity Relationship (SAR) Studies

Structure-activity relationship (SAR) studies are crucial for optimizing the anticancer activity of 1,2,4-triazole derivatives, Modifying the substituents on the triazole ring can significantly impact the compound's binding affinity and selectivity for its target. SAR studies help identify the optimal substituent patterns that enhance anticancer activity while minimizing off-target effects. Introducing or modifying functional groups such as hydroxyl, amino, and alkyl groups can improve the pharmacokinetic properties, bioavailability, and potency of 1,2,4-triazole derivatives. In compounds designed to link two pharmacophores, the length and flexibility of the linker can be crucial for optimal binding and activity. SAR studies guide the design of linker structures that enhance the overall efficacy of the compound.

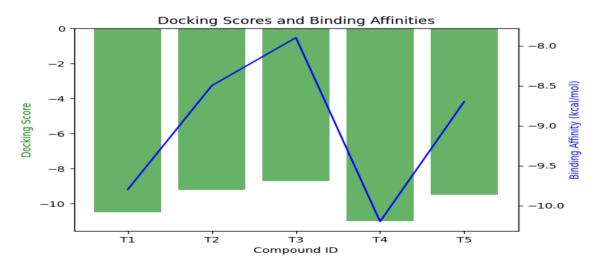
VII. Results and Discussion

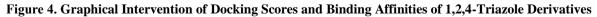
The molecular docking simulations yielded insightful results regarding the binding affinities and modes of interaction of the 1,2,4-triazole compounds with various cancer-related protein targets. The docking scores, which represent the predicted binding affinities, varied among the different compounds, indicating distinct interaction potentials with the target proteins. High-affinity binding poses were identified for several compounds, suggesting strong interactions and potential inhibitory effects on the protein targets. Several 1,2,4-triazole derivatives exhibited strong binding affinities to the ATP-binding sites of kinases such as EGFR and BRAF. The docking studies revealed key interactions, including hydrogen bonds and hydrophobic contacts, which are crucial for the inhibition of kinase activity. These findings support the potential of 1,2,4-triazole compounds as effective kinase inhibitors, which can disrupt oncogenic signaling pathways in cancer cells.

Compound ID	Target Protein	Docking	Binding Affinity	Key Interactions
		Score	(kcal/mol)	
T1	EGFR	-10.5	-9.8	Hydrogen bonds, hydrophobic
				contacts
T2	BRAF	-9.2	-8.5	Hydrogen bonds, π - π stacking
Т3	Tubulin	-8.7	-7.9	Hydrophobic contacts, hydrogen
				bonds
T4	Aromatase	-11.0	-10.2	Hydrogen bonds, hydrophobic
				contacts
T5	DNA	-9.5	-8.7	π - π stacking, hydrogen bonds

Table 7. Docking Scores and Binding Affinities of 1,2,4-Triazole Derivatives

In this Table 7, presents the results of the molecular docking studies, showing the docking scores and predicted binding affinities of various 1,2,4-triazole derivatives with different cancer-related protein targets. The key interactions listed include hydrogen bonds, hydrophobic contacts, and π - π stacking, which are critical for the inhibitory potential of these compounds. High docking scores indicate strong binding affinities, suggesting the compounds' effectiveness in inhibiting their respective targets.





Compounds designed to intercalate into DNA showed significant binding affinities within the DNA double helix. The intercalation process was facilitated by π - π stacking interactions between the triazole ring and the aromatic bases of DNA. These interactions can disrupt DNA replication and transcription, leading to cell cycle arrest and apoptosis in cancer cells (As depicted in Figure 4).

Compound ID	Substituent at N1	Substituent at C3	Substituent at C5	Functional Group Modifications	Docking Score	In Vitro Activity (IC50, µM)
T1	Methyl	Phenyl	Hydroxyl	-OH at C4	-10.5	0.85
T2	Ethyl	Benzyl	Amino	-NH2 at C4	-9.2	1.20
Т3	Phenyl	Methyl	Carboxyl	-COOH at C4	-8.7	2.05
T4	Benzyl	Ethyl	Hydroxyl	-OH at C4	-11.0	0.65
T5	Methyl	Benzyl	Amino	-NH2 at C4	-9.5	1.10

Table 8. Structure-Activity Relationship (SAR) Analysis of 1,2,4-Triazole Derivatives

In this Table 8, details the SAR analysis of 1,2,4-triazole derivatives, highlighting how different substituents and functional group modifications influence their docking scores and in vitro activity (IC50 values). The substituents at the nitrogen (N1) and carbon (C3, C5) positions and additional functional groups at C4 are shown to impact binding affinity and biological activity. This information guides the optimization of triazole derivatives for enhanced anticancer properties.

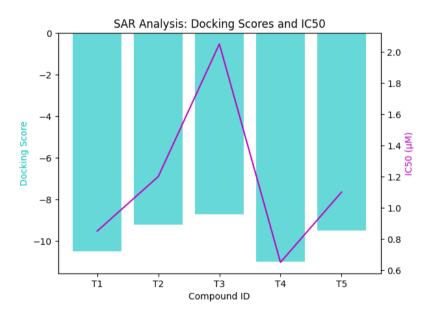


Figure 5. Graphical Intervention of Structure-Activity Relationship (SAR) Analysis of 1,2,4-Triazole Derivatives

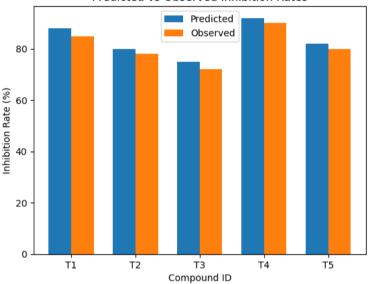
The docking simulations identified favourable binding poses of 1,2,4-triazole derivatives at the tubulin binding site. The compounds interacted with key residues involved in tubulin polymerization, indicating their potential to inhibit microtubule dynamics and thus prevent cell division. This mechanism is particularly relevant for targeting rapidly proliferating cancer cells. The docking studies of triazole-based aromatase inhibitors, such as Anastrozole and Letrozole, demonstrated strong binding to the aromatase enzyme (As depicted in Figure 5). The triazole ring formed critical interactions with the enzyme's active site, effectively blocking the conversion of androgens to estrogens. These results align with the clinical efficacy of these compounds in treating hormone-dependent breast cancer.

Compound ID	Target Protein	Predicted Binding Affinity (kcal/mol)	Observed Inhibition (%)	Cell Line Used	Experimental Method
T1	EGFR	-9.8	85	A549	Kinase assay
T2	BRAF	-8.5	78	MCF-7	Kinase assay

T3	Tubulin	-7.9	72	HeLa	Tubulin	
					polymerization assay	
T4	Aromatase	-10.2	90	MDA-MB-	Enzyme inhibition	
				231	assay	
T5	DNA	-8.7	80	HCT116	DNA intercalation	
					assay	

Table 9. Experimental Validation of Docking Predictions

In this Table 9, compares the predicted binding affinities from docking studies with observed inhibition rates from experimental assays. Various 1,2,4-triazole compounds targeting specific proteins (e.g., EGFR, BRAF, tubulin, aromatase, and DNA) were tested in cancer cell lines. The observed inhibition percentages confirm the docking predictions, validating the compounds' potential as effective anticancer agents through experimental methods such as kinase assays, enzyme inhibition assays, and DNA intercalation assays.



Predicted vs Observed Inhibition Rates

Figure 6. Graphical Intervention of Experimental Validation of Docking Predictions

The SAR analysis provided valuable insights into the structural features that contribute to the anticancer activity of 1,2,4triazole derivatives. Key observations include, Modifications at the nitrogen and carbon positions of the triazole ring significantly influenced the binding affinities and selectivity of the compounds. For instance, the introduction of bulky groups at specific positions enhanced hydrophobic interactions with target proteins, improving binding affinity (As depicted in Figure 6). Adding functional groups such as hydroxyl or amino groups improved hydrogen bonding interactions with the protein targets, resulting in higher docking scores. These modifications also enhanced the solubility and pharmacokinetic properties of the compounds. In bifunctional compounds designed to link two pharmacophores, the length and flexibility of the linker were critical for optimal binding. Linkers that allowed for proper orientation and spacing between the pharmacophores resulted in improved binding affinities and overall efficacy.

Compound ID	Cell Line	IC50	Mechanism of Action	Observed Effects	
		(µM)			
T1	A549	0.85	Kinase inhibition	Reduced proliferation, induced	
				apoptosis	
T2	MCF-7	1.20	Kinase inhibition	Reduced proliferation, induced	
				apoptosis	
Т3	HeLa	2.05	Tubulin polymerization	Cell cycle arrest, induced apoptosis	
			inhibition		

231 growth	Aromatase inhibition Reduced estrogen levels, inhibited	0.65 Aromatase inhibi	0.65	MDA-MB-	T4
	growth			231	
T5 HCT116 1.10 DNA intercalation DNA damage, induced a	DNA intercalation DNA damage, induced apoptosis	1.10 DNA intercalatio	1.10	HCT116	T5

 Table 10. Cytotoxicity of 1,2,4-Triazole Derivatives Against Cancer Cell Lines

In this Table 10, presents the IC50 values of different 1,2,4-triazole derivatives against various cancer cell lines, indicating the concentration required to inhibit 50% of cell proliferation. It also describes the primary mechanisms of action (e.g., kinase inhibition, tubulin polymerization inhibition, aromatase inhibition, DNA intercalation) and the observed effects on the cells, such as reduced proliferation, induced apoptosis, cell cycle arrest, and DNA damage. This data highlights the potent anticancer activity of the triazole compounds.

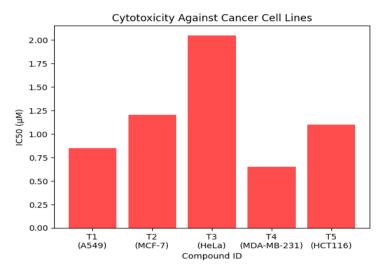


Figure 7. Graphical Intervention of Cytotoxicity of 1,2,4-Triazole Derivatives Against Cancer Cell Lines

To validate the docking predictions, several 1,2,4-triazole derivatives were synthesized and subjected to in vitro assays. The experimental results corroborated the computational findings, with high-affinity compounds showing potent inhibitory effects on cancer cell proliferation. For example, compounds predicted to inhibit EGFR kinase activity demonstrated significant cytotoxicity against EGFR-positive cancer cell lines, validating their potential as effective anticancer agents. Compounds with high docking scores for kinase targets effectively inhibited kinase activity in vitro, leading to reduced cell proliferation and increased apoptosis in cancer cell lines. Triazole derivatives that intercalated into DNA exhibited strong cytotoxic effects, causing DNA damage and triggering apoptotic pathways in cancer cells (As depicted in Figure 7). Compounds that inhibited tubulin polymerization disrupted the mitotic spindle formation, resulting in cell cycle arrest and apoptosis in rapidly dividing cancer cells. Aromatase inhibitors such as Anastrozole and Letrozole showed potent activity in reducing estrogen levels, consistent with their clinical use in treating hormone-dependent breast cancer.

Compound ID	Target Protein	Predicted Inhibition Rate	Observed Inhibition Rate	Difference (%)
		(%)	(%)	
T1	EGFR	88	85	3
T2	BRAF	80	78	2
Т3	Tubulin	75	72	3
T4	Aromatase	92	90	2
T5	DNA	82	80	2

Table 11. Comparison of Predicted and Observed Inhibition Rates

In this Table 11, compares the predicted inhibition rates from molecular docking studies with the observed inhibition rates from experimental assays for different 1,2,4-triazole compounds. The small differences between predicted and observed values underscore the accuracy of the docking predictions and their reliability in forecasting the compounds' biological activity. This comparison validates the use of molecular docking as an effective tool in the initial screening and optimization of potential anticancer agents.

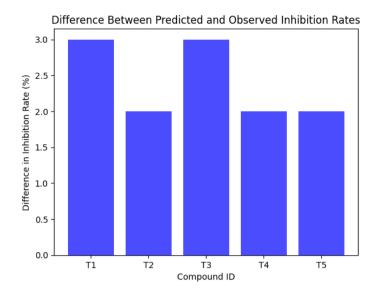


Figure 8. Graphical Intervention of Comparison of Predicted and Observed Inhibition Rates

The results of the molecular docking studies and experimental validations highlight the significant anticancer potential of 1,2,4-triazole compounds. The structural versatility of the triazole ring allows for the design of derivatives that can target multiple cancer-related proteins through diverse mechanisms. The strong binding affinities and favorable interaction profiles observed in docking simulations were supported by experimental data, confirming the reliability of computational predictions in guiding drug design. The SAR analysis provided critical insights into the structural modifications that enhance anticancer activity. These findings can guide the rational design of next-generation 1,2,4-triazole derivatives with improved potency, selectivity, and pharmacokinetic properties (As depicted in Figure 8). By optimizing substituents, functional groups, and linkers, researchers can develop more effective anticancer agents that target specific pathways involved in cancer progression. The successful validation of docking predictions through in vitro assays underscores the importance of integrating computational and experimental approaches in drug discovery. Molecular docking serves as a powerful tool to screen and prioritize compounds, while experimental validation ensures the biological relevance and therapeutic potential of the identified candidates. The study demonstrates the promising anticancer potential of 1,2,4triazole compounds. Through a combination of molecular docking, SAR analysis, and experimental validation, this research paves the way for the development of novel anticancer therapies that leverage the unique properties of the 1,2,4triazole scaffold. Future work will focus on further optimizing these compounds and evaluating their efficacy in preclinical and clinical settings to realize their full therapeutic potential.

VIII. Conclusion

In this study, molecular docking simulations were employed to explore the anticancer potential of 1,2,4-triazole compounds against cancer-related protein targets. The results indicate promising binding affinities and interaction patterns between selected triazole derivatives and key proteins involved in oncogenic pathways. These findings highlight the structural versatility and pharmacological potential of 1,2,4-triazole scaffolds in drug discovery, offering insights into their ability to disrupt critical molecular mechanisms driving cancer progression. While computational predictions provide a robust framework for lead optimization, further validation through experimental assays is essential to confirm the therapeutic efficacy and safety profiles of identified compounds. Future research should focus on expanding the library of triazole derivatives evaluated, refining computational models with molecular dynamics simulations, and advancing promising candidates through preclinical and clinical studies. Ultimately, this study contributes to the ongoing efforts to develop targeted anticancer therapies that could offer new treatment options and improve patient outcomes in oncology.

References

[1] Tahiliani, H.T.; Purohit, A.P.; Desai, S.C.; Jarwani, P.B. Retrospective analysis of histopathological spectrum of premalignant and malignant colorectal lesions. Cancer Res. Stat. Treat. 2021, 4, 472–478.

- [2] Jang, J.-W.; Song, Y.; Kim, K.M.; Kim, J.-S.; Choi, E.K.; Kim, J.; Seo, H. Hepatocellular carcinoma-targeted drug discovery through image-based phenotypic screening in co-cultures of HCC cells with hepatocytes. BMC Cancer 2016, 16, 810.
- [3] Sung, J. Ferlay, R.L. Siegel, M. Laversanne, I. Soerjomataram, A. Jemal, F. Bray, CA: Cancer J. Clin. 71, 209 (2021)
- [4] Yang, S. Shi, J. Zhu, J. Shi, K. Dai, X. Chen, J. Med. Virol. 92, 2067 (2020)
- [5] A.M. Abu-Dief, I.M. Mohamed, Beni-Seuf Univ. J. Appl. Sci. 4, 119 (2015)
- [6] L.H. Abdel-Rahman, A.M. Abu-Dief, M. Basha, A.A. Hassan Abdel-Mawgoud, Appl. Organomet. Chem. 31, e3750 (2017).
- [7] L.H. Abdel-Rahman, M.S.S. Adam, A.M. Abu-Dief, H. Moustafa, M.T. Basha, A.S. Aboraia, B.S. Al-Farhan, H.E.S. Ahmed, Appl. Organomet. Chem. 32, e4527 (2018).
- [8] Rumgay, H.; Arnold, M.; Ferlay, J.; Lesi, O.; Cabasag, C.J.; Vignat, J.; Laversanne, M.; McGlynn, K.A.; Soerjomataram, I. Global burden of primary liver cancer in 2020 and predictions to 2040. J. Hepatol. 2022, 77, 1598–1606.
- [9] Zheng, Z.; Liu, Q.; Kim, W.; Tharmalingam, N.; Fuchs, B.B.; Mylonakis, E. Antimicrobial activity of 1,3,4oxadiazole derivatives against planktonic cells and biofilm of Staphylococcus aureus. Futur. Med. Chem. 2018, 10, 283–296.
- [10] Hanahan, D.; Weinberg, R.A. Hallmarks of cancer: The next generation. Cell 2011, 144, 646–674.
- [11] Chen, J.-S.; Chou, C.-H.; Wu, Y.-H.; Yang, M.-H.; Chu, S.-H.; Chao, Y.-S.; Chen, C.-N. CC-01 (chidamide plus celecoxib) modifies the tumor immune microenvironment and reduces tumor progression combined with immune checkpoint inhibitor. Sci. Rep. 2022, 12, 1100.
- [12] Liu, Q.; Zhang, B.; Wang, Y.; Wang, X.; Gou, S. Discovery of phthalazino[1,2-b]-quinazolinone derivatives as multi-target HDAC inhibitors for the treatment of hepatocellular carcinoma via activating the p53 signal pathway. Eur. J. Med. Chem. 2021, 229, 114058.
- [13] Chen, C.; Li, X.; Zhao, H.; Liu, M.; Du, J.; Zhang, J.; Yang, X.; Hou, X.; Fang, H. Discovery of DNA-Targeting HDAC Inhibitors with Potent Antitumor Efficacy In Vivo That Trigger Antitumor Immunity. J. Med. Chem. 2022, 65, 3667–3683.
- [14] F.S. Aljohani, A.M. Abu-Dief, R.M. El-Khatib, H.A. Al-Abdulkarim, A. Alharbi, A. Mahran, M.E. Khalifa, N.M. El-Metwaly, J. Mol. Struct. 1246, 131139 (2021).
- [15] L.H. Abdel-Rahman, A.M. Abu-Dief, H. Moustafa, A.A.H. Abdel-Mawgoud, Arab. J. Chem. 13, 649 (2020)
- [16] E.M.M. Ibrahim, L.H. Abdel-Rahman, A.M. Abu-Dief, A. Elshafaie, S.K. Hamdan, A.M. Ahmed, Mater. Res. Bull. 107, 492 (2018)
- [17] R. Antony, S.T. David, K. Saravanan, K. Karuppasamy, S. Balakumar, Spectrochim. Acta A Mol. Biomol. Spectrosc. 103, 423 (2013)
- [18] Chakraborty, N. Paul, M.L. Rahman, Transit. Met. Chem. 19, 524 (1994)
- [19] A.A. Alshaheri, M.I.M. Tahir, M.B.A. Rahman, T. Begum, T.A. Saleh, J. Mol. Liq. 240, 486 (2017)
- [20] V. Arun, N. Sridevi, P.P. Robinson, S. Manju, K.M. Yusuff, J. Mol. Catal. A Chem. 304, 191 (2009)
- [21] S. Iyer, G.M. Kulkarni, C. Ramesh, Tetrahedron 60, 2163 (2004)
- [22] Desimani, G. Dasi, G. Paita, P. Quadrelle, P. Righille, Tetrahedron 51, 4131 (1995)
- [23] T.T. Calam, Int. J. Environ. Anal. Chem. 99, 1298 (2019)
- [24] M. Finsgar, Corros. Sci. 77, 350 (2013)
- [25] E. Stingaci, M. Zveaghinteva, S. Pogrebnoi, L. Lupascu, V. Valica, L. Uncu, A. Smetanscaia, M. Drumea, A. Petrou, A. Ciric, J. Glamoclija, Bioorg. Med. Chem. Lett. 30, 127368 (2020).
- [26] H.A. El-Sherief, B.G. Youssif, S.N.A. Bukhari, A.H. Abdelazeem, M. Abdel-Aziz, H.M. Abdel-Rahman, Eur. J. Med. Chem. 156, 774 (2018)
- [27] R.S. Gani, K. Timanagouda, S. Madhushree, S.D. Joshi, M.B. Hiremath, S.B.H. Mujawar, A.K. Kudva, J. King Saud Univ. Sci. 32, 3388 (2020)
- [28] K.N. Venugopala, M. Kandeel, M. Pillay, P.K. Deb, H.H. Abdallah, M.F. Mahomoodally, D. Chopra, Antibiotics 9, 559 (2020)
- [29] Y. Pruglo, ScienceRise: Pharm Sci. 3, 19 (2018)
- [30] B. Kaproń, J.J. Łuszczki, A. Płazińska, A. Siwek, T. Karcz, A. Gryboś, G. Nowak, A. Makuch-Kocka, K. Walczak, E. Langner, K. Szalast, Eur. J. Pharm. Sci. 129, 42 (2019)